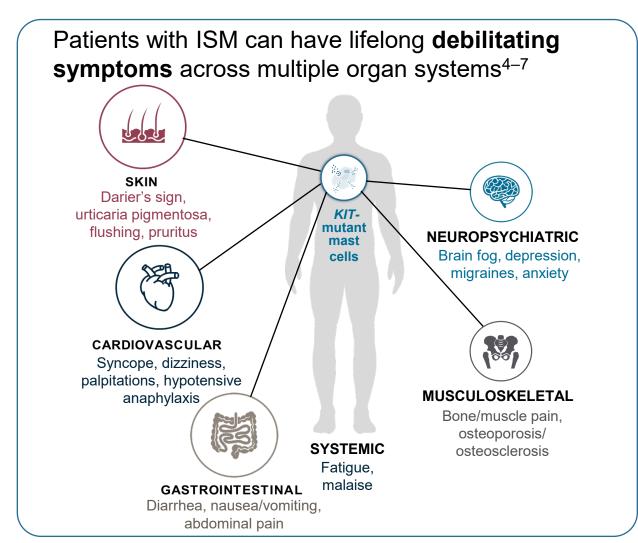
Responses to Avapritinib in Patients Without Detectable *KIT*Mutations by ddPCR in Peripheral Blood Highlight Diagnostic Challenges and Opportunities in Indolent Systemic Mastocytosis

Thanai Pongdee,¹ Pankit Vachhani,² Tsewang Tashi,³ Ivan Alvarez-Twose,⁴ Patrizia Bonadonna,⁵ Deepti H. Radia,⁶ Hanneke Oude Elberink,⁷ Jason Gotlib,⁸ Sonia Cerquozzi,⁹ Vito Sabato,¹⁰ Ingunn Dybedal,¹¹ Massimo Triggiani,¹² Cristina Bulai Livideanu,¹³ Karin Hartmann,^{14,15} Stephen Oh,¹⁶ Prithviraj Bose,¹⁷ Ruchi Desai,¹⁸ Phillipe Schafhausen,¹⁹ Marcus Maurer,^{20,21} Frank Siebenhaar,^{20,21} Tracy I. George,^{22,23} Sigurd Broesby-Olsen,²⁴ Guang Yang,²⁵ Hui-Min Lin,²⁵ Mark Rosenzweig,²⁵ Ben Lampson,²⁵ Rachel L. Erlich,²⁵ Cem Akin²⁶

¹Division of Allergic Diseases, Mayo Clinic, Rochester, MN, USA; ²Department of Medicine, University of Alabama at Birmingham, Birmingham, AL, USA; ³Huntsman Cancer Institute, University of Utah, Salt Lake City, UT, USA; ⁴Institute of Mastocytosis Studies of Castilla-La Mancha, Virgen del Valle Hospital, Toledo, Spain; ⁵Allergy Unit and Asthma Center, Azienda Ospedaliera Universitaria Integrata di Verona, Verona, Italy; ⁰Guy's & St Thomas' NHS Foundation Trust, London, United Kingdom; ¹Department of Allergology, University Medical Center, Groningen Research Institute Asthma and COPD, University of Groningen, Groningen, The Netherlands; ⁰Stanford Cancer Institute/Stanford University School of Medicine, Stanford, CA, USA; ⁰Department of Medicine, Cumming School of Medicine, University of Calgary, Calgary, Alberta, Canada; ¹¹Department of Immunology, Allergology and Rheumatology, University of Antwerp, and Antwerp University Hospital, Antwerp, Belgium; ¹¹Department of Hematology and Pharmacology, Oslo University Hospital, Oslo, Norway; ¹²Division of Allergy and Clinical Immunology, University of Salerno, Italy; ¹³Department of Dermatology, Expert Center of Mastocytosis (CEREMAST), Toulouse University Hospital, Toulouse, France; ¹⁴Division of Allergy, Department of Dermatology, University Hospital Basel and University of Basel, Basel, Switzerland; ¹⁵Stepartment of Bormatology, University Hospital Basel and University of Basel, Switzerland; ¹⁵Stepartment of Bormatology, University of Cancer Center at Barnes-Jewish Hospital and Washington University School of Medicine, Saint Louis, MO, USA; ¹³Department of Leukemia, Division of Cancer Medicine, The University of Texas MD Anderson Cancer Center, Houston, TX, USA; ¹⁰Division of Hematology and Oncology, Virginia Commonwealth University, Massey Comprehensive Cancer Center, Richmond, VA, USA; ¹³Department of Oncology, Hematology, and Bone Marrow Transplantation with Section of Pneumology, University Medical Center Hamburg-Eppendorf, Hamburg, Germany; ²¹Fraunhofer Institut

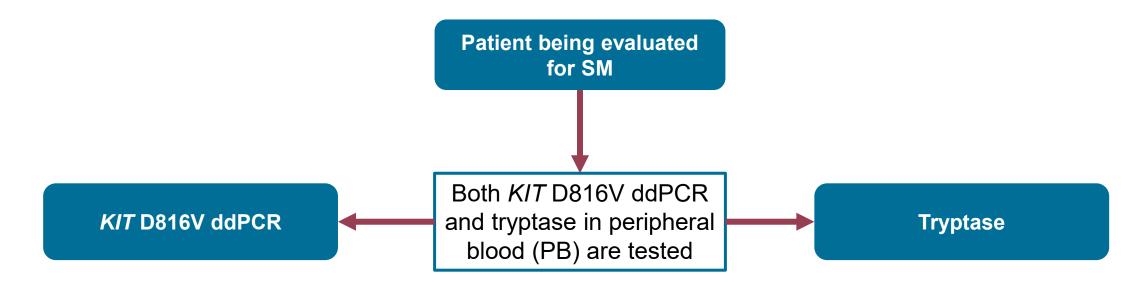
Indolent systemic mastocytosis: A *KIT* D816V mutation–driven disease with substantial impact on quality of life

- Indolent systemic mastocytosis (ISM), the most common subtype of systemic mastocytosis (SM), is driven by aberrant mast cells carrying a KIT D816V mutation in ~95% of cases^{1–3}
- The diagnosis of SM is made according to a set of criteria defined by expert consensus^{8–10}
- One of the diagnostic criteria is demonstrating the presence of a KIT mutation
 - KIT mutations can be difficult to detect in blood due to low levels of circulating KIT-mutant cells in ISM
 - SM cannot be ruled out if ddPCR does not detect a mutation in blood: a bone marrow biopsy is still required if suspicion is high



ISM, indolent systemic mastocytosis.

Peripheral blood testing in a patient with suspected SM has advantages but could be improved



Advantages

- · Samples easily collected (blood draw), tested, and interpreted
- Droplet digital polymerase chain reaction (ddPCR) has the highest sensitivity for KIT D816V mutation out of the current clinically available tests (much higher than next generation sequencing)¹¹
- Presence of KIT activating mutation is a minor diagnostic criterion for SM

Disadvantages

- Imperfect sensitivity: only 85% of patients with ISM were detected in the PIONEER study¹²
- Cannot be used as a "rule-out" test, bone marrow biopsy is still required if suspicion is high

Advantages

- · Samples easily collected and tested (blood draw)
- Tryptase levels >20 ng/mL is a minor diagnostic criterion for SM⁹

Disadvantages

- Interpretation can be challenging, hereditary alpha tryptasemia status is needed
- Poor sensitivity, as up to 30% of patients with SM may have tryptase <20 ng/mL¹³
- Cannot rule out SM even if tryptase levels <20 ng/mL

Detection methods for *KIT* mutations in SM vary in sensitivity: ddPCR is the current gold standard and more sensitive than NGS

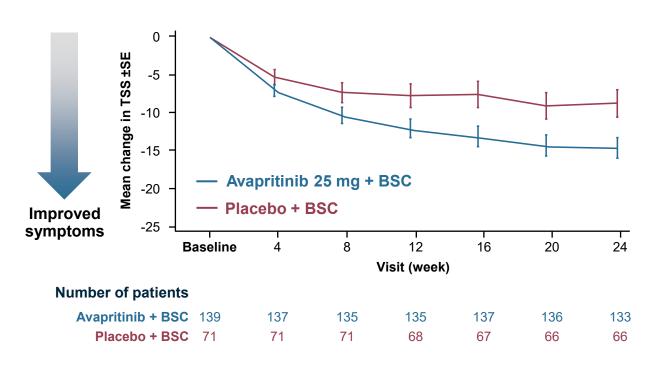
Technology	Assay status	LOD for <i>KIT</i> D816V mutations	KIT mutations that can be detected	Sample input	Useful for ISM diagnosis?
NGS	Commercial use	5% ¹¹	Multiple exon 17 mutations	Isolated DNA from blood or bone marrow aspirate	Only detects <i>KIT</i> D816V in ~30% of patients ¹⁴
ddPCR		0.022%12	D816V only		Current gold standard, positive in ~85% of patients ¹²
Duplex sequencing	Research use	0.0013% ¹⁶	Multiple exon 17 mutations		17x more sensitive than ddPCR

To determine whether ultra-sensitive duplex sequencing facilitates detection of more *KIT* mutations, we evaluated its use on clinical trial samples from patients with verified ISM who had no detectable *KIT* mutation by ddPCR

The cohort of patients enrolled in the PIONEER trial of avapritinib represents an opportunity to better understand ISM

- PIONEER (NCT03731260) is a double-blind, placebo-controlled trial of avapritinib in patients with ISM¹²
- Avapritinib potently and selectively inhibits
 KIT D816V¹⁷
- In PIONEER, avapritinib significantly improved total symptom score (TSS) as assessed by the ISM-Symptom Assessment Form^a (ISM-SAF),¹² leading to approval for adults with ISM in the USA and Europe^{18,19}
- PIONEER required ddPCR testing for KIT D816V mutations in all patients at time of enrollment

Mean change in ISM-SAF TSS over time



Use of patient samples from PIONEER allowed avapritinib response assessment in patients who did not have detectable *KIT* D816V mutations by ddPCR

Patients who enrolled in PIONEER had peripheral blood testing for *KIT* mutations at screening and subdivided into groups

ddPCR

Sequencing

Duplex

 Patients who had no detectable KIT D816V in PB by ddPCR were further tested with duplex sequencing

 Of 37 patients with no detectable KIT D816V by ddPCR, 26 had activating KIT mutations detectable by duplex sequencing

 Combining results from clinical ddPCR testing and research duplex sequencing, 96% of patients from PIONEER had detectable activating KIT mutations

KIT mutations No detectable *KIT* 246 patients were detected mutations tested at screening Overall 235/246 **Overall 11/246** 209/246 (85%) had 37/246 (15%) had detectable KIT no detectable KIT **D816V mutations D816V mutations** 23/37 (62%) had 11/37 (30%) had no detectable KIT detectable KIT **D816V mutations** D816V or other exon 17 KIT 3/37 (8%) had mutations detectable non-**D816V exon 17 KIT** mutations only

Patients with *KIT* mutations detectable only by duplex sequencing had a lower baseline disease burden

Characteristic	KIT mutation detectable by ddPCR (n=209)	KIT mutation not detectable by ddPCR and detectable by duplex sequencing (n=26)	P-value
Age, years (range)	51 (18–79)	48 (31–64)	0.24
Female, %	153 (73)	20 (77)	0.82
Median baseline serum tryptase, ng/mL (range)	43.1 (4.2–590.4)	23.4 (3.6–250.4)	<0.01
Median BM MC, % (range)	10 (1.0–60.0)	5.0 (1.0–40.0)	<0.001
Median KIT D816V VAF, % (range)	0.51 (0.02–41.3)	0.0068 (0.0013–0.0261)	<0.0001

Duplex sequencing also successfully identified non-canonical *KIT* mutations that cannot be detected by ddPCR

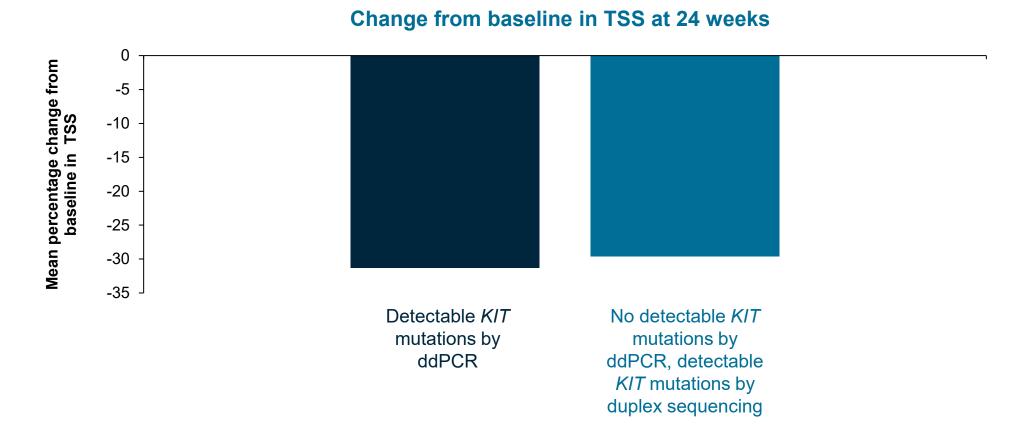
- A total of 21/26 patients had a detectable lone KIT D816V mutation
- Other *KIT* mutations were detected in 5/26 patients, including:
 - Patients (n=2) with dual mutations in KIT (D816I+D816V, C788Y+D816V)
 - Patients (n=3) with lone non-D816V KIT activating mutations (D816I, D816Y; VAF 0.0075–4.5%)

Patients with non-D816V KIT mutations detected by duplex sequencing

Age, years	Gender	Mutations detected	Median <i>KIT</i> mutation VAF, % by duplex sequencing	Avapritinib sensitivity in vitro (IC ₅₀ <1 nm) ¹⁸
63	Female	D816I/D816V	0.0013/0.0026	Yes/Yes
33	Female	D816I	0.7820	Yes
52	Male	D816Y	4.4781	Yes
51	Female	D816Y	0.0075	Yes
31	Female	C788Y/D816V	0.0041/0.0037	ND/Yes

Similar improvements were seen in mean percentage change from baseline in TSS irrespective of the test used to detect *KIT* mutations

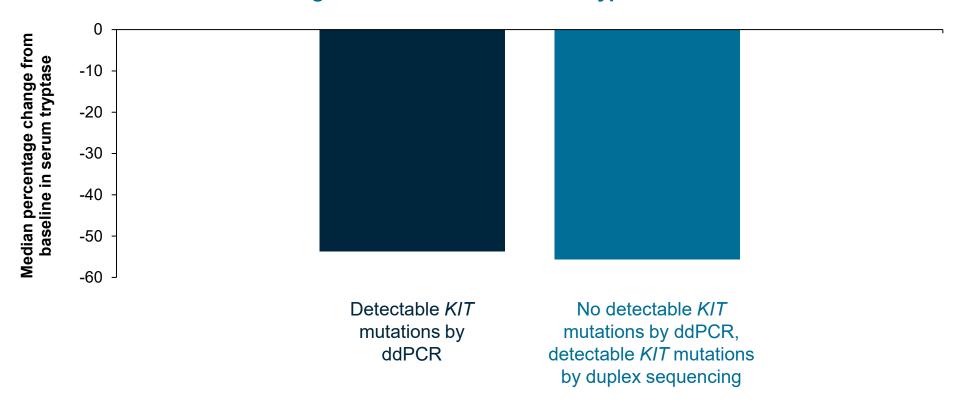
• After 24 weeks of therapy, improvements were seen for avapritinib-treated patients^a in mean percentage change from baseline in TSS whether *KIT* mutations were detected by ddPCR (n=194) or by duplex sequencing (n=22)



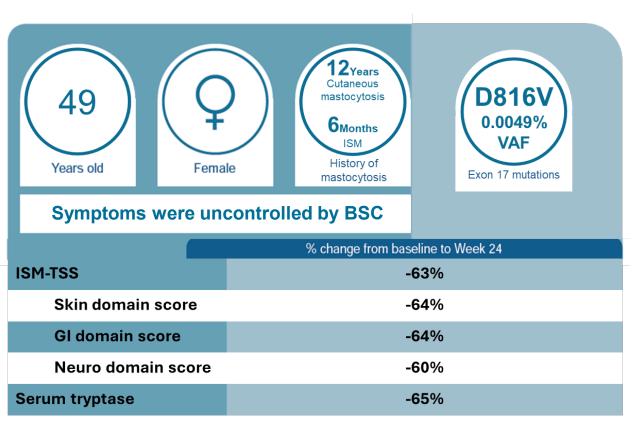
Similar median percentage change from baseline in serum tryptase levels in avapritinib-treated patients at 24 weeks by *KIT* mutational status

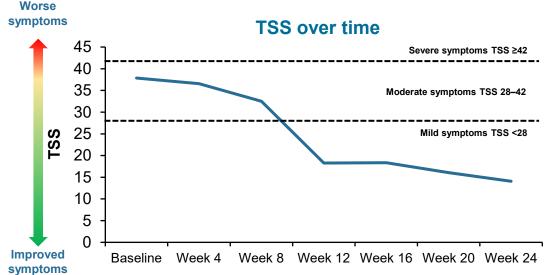
After 24 weeks of avapritinib treatment, improvements were seen in tryptase percentage change from baseline in patients^a whether KIT mutations were detected by ddPCR (n=194) or by duplex sequencing only (n=22)

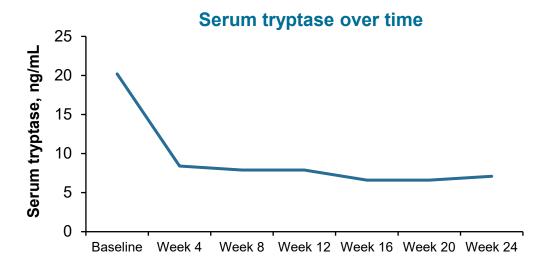
Change from baseline in serum tryptase at 24 weeks



Informative case study: Effectiveness of avapritinib in a patient with a *KIT* D816V mutation below the VAF detection threshold of ddPCR







Conclusions

- Due to the rarity of circulating mutant cells in PB in ISM, more sensitive assays are needed to aid clinicians in identifying *KIT* D816V mutations, an important minor diagnostic criterion
- While serum tryptase and ddPCR testing for *KIT* D816V in PB are important tests in the work-up of suspected SM, the possibility of SM cannot be ruled out when *KIT* D816V is not detected
- The combination of ddPCR testing and ultra-sensitive duplex sequencing was able to identify an activating exon 17 *KIT* mutation in the blood of 96% of patients with ISM in PIONEER
 - We found that 70% of patients with undetectable KIT D816V by ddPCR had activating KIT mutations detected by duplex sequencing
- Avapritinib can effectively reduce symptoms even in patients who do not have detectable KIT D816V by ddPCR
- Bone marrow biopsy, including ddPCR of the bone marrow aspirate sample for *KIT* D816V, is still the standard-of-care for evaluating SM and should be performed if SM is suspected

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Conflicts of interest and disclosures

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